



# Gene therapy and stem cells in the treatment of congenital diseases

# Paula Rubya de Souza Câmara

# ABSTRACT

Department of Medicine, Potiguar University, Natal, Rio Grande do Norte, Brazil

Address for correspondence: Paula Rubya de Souza Câmara, Av. Senador Salgado Filho, 1610, Natal, Rio Grande do Norte 59056-000, Brazil. Phone: +55-84-3215-1273, Fax: +55-84-3215-1200, E-mail: aluapcamara@ hotmail.com

Received: November 03, 2014 Accepted: January 28, 2015 Published: March 07, 2015 Several congenital diseases are particularly attractive candidates for intervention using gene therapy since the underlying molecular bases for most of the monogenic disorders are well-understood. Transplantation of *ex vivo* genetically modified stem cells has also shown promise. Although all of these systems are meritorious and worthy of continued investigation, this mini-review article focused on the platforms that have received the most attention and that are maturing in the clinical setting; in particular, the potential of *in vivo* gene therapy and human-induced pluripotent stem cells. Studies of apparently disparate diseases that are presumably linked through shared metabolic pathways are likely to provide greater insights into the biology of the diseases. This and other opportunities for exchange will hopefully foster acceleration in the development of new and innovative therapies for these devastating diseases.

KEY WORDS: Congenital diseases, frozen embryos, gene therapy, pluripotency, stem cells

# INTRODUCTION

Congenital diseases are responsible for over a third of all pediatric hospital admissions. Advances in prenatal screening and molecular diagnosis have allowed the detection of many life-threatening genetic diseases early in gestation. The options of preemptive treatment of congenital diseases in utero by gene therapy or stem cell changes the perspective of congenital diseases since it may avoid the need for postnatal treatment and reduce future costs.

# **GENE THERAPY**

Gene therapy is promising in the treatment of many congenital diseases. With all the hoopla surrounding the Human Genome Project, it is understandable that people would entertain high hopes for the advancement of gene therapy. It falls into three groups: (1) replacing a defective or bad adaptive gene that's responsible for some monogenic disease, (2) altering or killing an aberrant cell (e.g. infected by HIV or cancerous) and (3) inducing production of a therapeutic protein. Initially, gene therapy focused on the first group, but most current research focuses on the other two. Whatever the application, numerous hurdles stand in the way of developing a successful gene therapy. These obstacles include identifying an appropriate target for gene therapy such as getting a therapeutic transgene into the right cells (and only those cells) in the right amount; delivering the transgene with a vector that does not trigger an

58

immune response or, in the case of certain viral vectors, revert to a pathogenic form; providing the appropriate regulatory elements for turning the gene on and off at the correct time; keeping the transgene in the target cell long enough for it to do its job; and keeping the transgene from causing damage elsewhere [1-6]. The advances in gene therapy hold significant promise for the treatment of ophthalmic conditions such as heritable diseases of the retina [7-9], endocrinology [10], rheumatic diseases [11-13], Alzheimer's disease [14], diseases of the gastrointestinal tract [15-22], therapy of cancer [23-27] and neurological disorders [28,29]. In addition, diseases like inborn errors of metabolism (include the diseases resulting from enzyme defects in biochemical reactions due to genetic mutations), and human peroxisomal disorders (caused by peroxisomal ABC half-transporters mutation which is localized in the peroxisomal membrane) could be candidate to gene therapy [30-33].

In addition, artificial chromosomes (ACs) are highly promising vectors for use in gene therapy applications. They are able to maintain expression of genomic-sized exogenous transgenes within target cells, without integrating into the host genome. The recent developments in AC technology present improved methods for the production, purification, delivery, and natural transgene expression of genomic sized loci. These technologies are all steps forward in alleviating problems associated with synthetically produced cDNA. In addition, AC technology is proving to be highly compatible with stem-cell research. With further development, ACs could be used to improve the efficacy of gene therapy by providing physiologically appropriate expression of transgenes *in vivo* in target tissues [34]. Moreover, carbon nanotubes have been proposed and are actively being explored as innovative multipurpose carriers for biomolecules and diagnostic applications. They are used in the controlled release of drugs as well as delivery of genetic material such as DNA, genes, and antibodies [35-37].

Ethical and political issues need to be addressed, but, over the long-term, the future of drug therapy could be gene therapy. Then, this article focused on the potential of *in vivo* gene therapy and alternative sources of pluripotency as perspectives in the treatment of congenital diseases.

#### ALTERNATIVE SOURCES OF PLURIPOTENCY

It has recently been shown that the first cleavage plane of the mouse zygote defines the border between the embryonic and abembryonic parts of the blastocyst and that this border correlates with the sperm entry position (SEP). Plusa *et al.* [38] developed a means of fluorescently labeling sperm that can record the exact site of its penetration when the label transfers to the egg surface. This approach indicates that the SEP marks the first cleavage in the great majority (88%) of embryos, e.g. life.

In the same way, Takahashi et al. [39] and Yu et al. [40] suggested alternative sources of pluripotency. They demonstrated that expression of four specific transcription factors (Oct4, Sox2, Klf4, and c-Myc) gives adult human fibroblasts many of the characteristics of human embryonic stem cells (hESCs). Refinements of this procedure will make it possible to produce pluripotent human cell lines without the use of an embryo [41-46]. In addition, human induced pluripotent stem (iPS) cells obtained by reprogramming technology are a source of great hope, not in terms of applications in regenerative medicine, such as cell transplantation therapy, but also for modeling human diseases and new drug development. In particular, the production of iPS cells from the somatic cells of patients with intractable diseases and their subsequent differentiation into cells at affected sites (e.g., neurons, cardiomyocytes, hepatocytes, and myocytes) has permitted the in vitro construction of disease models that contain patient-specific genetic information [47-95]. For example, disease-specific iPS cells have been established from patients with neuropsychiatric disorders, including schizophrenia and autism, as well as from those with neurodegenerative diseases, including Parkinson's disease and Alzheimer's disease [96-102].

Finally, stem cells have the capability to proliferate and differentiate into various cells of the body. Few stem cell sources have been approved for transplantation; among them are the hematopoietic progenitor cells which are progenitors of the myeloid and erythroid lineage in the hematopoietic system that continually provides mature blood cells during the lifespan of the individual. These well-characterized stem cells are clinically relevant in the treatment of diseases such as breast cancer, leukemias, and congenital immunodeficiencies. In addition, the investigation of mesenchymal stem cells secretome is accumulating continuously increasing interest given the potential use of these cells in regenerative medicine [103-107].

De Coppi et al. [108] reported the isolation of human and rodent amniotic fluid-derived stem (AFS) cells that express embryonic and adult stem cell markers. Undifferentiated AFS cells expand extensively without feeders, double in 36 h and are not tumorigenic. Lines maintained for over 250 population doublings retained long telomeres and a normal karyotype. AFS cells are broadly multipotent. Clonal human lines verified by retroviral marking were induced to differentiate into cell types representing each embryonic germ layer, including cells of adipogenic, osteogenic, myogenic, endothelial, neuronal, and hepatic lineages. Differentiated cell derived from human AFS cells are displaying specialized functions which include neuronal lineage cells secreting the neurotransmitter L-glutamate or expressing G-protein-gated inwardly rectifying potassium channels, hepatic lineage cells producing urea, and osteogenic lineage cells forming tissue-engineered bone [109-116].

Despite many advances in hESC technology; the ethical dilemma involving the destruction of a human embryo is an important factor limiting the development of hESC based clinical therapies [117]. The application of embryo freezing to human *in-vitro* fertilization (IVF) has revolutionized its clinical practice and helped to convert IVF from an experimental procedure to the widespread practice. In Australia, more than 7000 babies have been born following the transfer of frozen-thawed embryos [118,119].

It is worth considering how couples can be encouraged to donate rather than discard their surplus frozen embryos. At present, there are numerous frozen embryos ready for donation. Moreover, it has been speculated that the primary means by which reactive oxygen species reduce the fertility of semen subjected to refrigeration or long-term liquid storage is its impact on sperm DNA integrity [120,121]. An educational program around the world on relevant legal, social, and clinical issues may facilitate this, and therefore, give an ethic destination to the human embryo frozen.

#### Author contribution

Paula RS Câmara wrote the paper and reviewed it.

#### REFERENCES

- 1. McCain J. The future of gene therapy. Biotechnol Healthc 2005;2:52-60.
- 2. Malik P, Arumugam PI. Gene Therapy for beta-thalassemia. Hematology Am Soc Hematol Educ Program 2005:45-50.
- Löscher W, Gernert M, Heinemann U. Cell and gene therapies in epilepsy-promising avenues or blind alleys? Trends Neurosci 2008;31:62-73.
- Simonato M. Gene therapy for epilepsy. Epilepsy Behav 2014;38:125-30.
- Macpherson JL, Rasko JE. Clinical potential of gene therapy: towards meeting the demand. Intern Med J 2014;44:224-33.
- High KH, Nathwani A, Spencer T, Lillicrap D. Current status of haemophilia gene therapy. Haemophilia 2014;20 Suppl 4:43-9.
- Al-Saikhan FI. The gene therapy revolution in ophthalmology. Saudi J Ophthalmol 2013;27:107-11.

- Bible E. Sensory systems: Promising results in a gene therapy trial for retinal disease. Nat Rev Neurol 2014;10:123.
- MacLaren RE, Groppe M, Barnard AR, Cottriall CL, Tolmachova T, Seymour L, *et al*. Retinal gene therapy in patients with choroideremia: initial findings from a phase 1/2 clinical trial. Lancet 2014;383:1129-37.
- Barzon L, Bonaguro R, Palù G, Boscaro M. New perspectives for gene therapy in endocrinology. Eur J Endocrinol 2000;143:447-66.
- Evans CH, Ghivizzani SC, Kang R, Muzzonigro T, Wasko MC, Herndon JH, *et al.* Gene therapy for rheumatic diseases. Arthritis Rheum 1999;42:1-16.
- 12. Evans CH, Ghivizzani SC, Robbins PD. Gene therapy of the rheumatic diseases: 1998 to 2008. Arthritis Res Ther 2009;11:209.
- 13. Evans CH, Ghivizzani SC, Robbins PD. Arthritis gene therapy and its tortuous path into the clinic. Transl Res 2013;161:205-16.
- 14. Iwata N, Ozawa K, Saido TC. Gene therapy for Alzheimer's disease. Nihon Rinsho 2005;63:394-400.
- Kaiser R, Thiel E, Kreuser ED. Human gene therapy in gastrointestinal diseases: *In vivo* and *in vitro* approaches. Recent Results Cancer Res 1996;142:51-61.
- Blum HE, Wieland S, von Weizsäcker F. Gene therapy: basic concepts and applications in gastrointestinal diseases. Digestion 1997;58:87-97.
- Forbes SJ, Hodgson HJ. Review article: gene therapy in gastroenterology and hepatology. Aliment Pharmacol Ther 1997;11:823-36.
- Gottschalk U, Chan S. Somatic gene therapy. Present situation and future perspective. Arzneimittelforschung 1998;48:1111-20.
- Camilleri M. Enteric nervous system disorders: genetic and molecular insights for the neurogastroenterologist. Neurogastroenterol Motil 2001;13:277-95.
- 20. Flotte TR, Laube BL. Gene therapy in cystic fibrosis. Chest 2001;120 3 Suppl:124S-31S.
- Prieto J, Herraiz M, Sangro B, Qian C, Mazzolini G, Melero I, *et al.* The promise of gene therapy in gastrointestinal and liver diseases. Gut 2003;52 Suppl 2:ii49-54.
- Prieto J, Qian C, Hernandez-Alcoceba R, Gonzalez-Aseguinolaza G, Mazzolini G, Sangro B, *et al*. Gene therapy of liver diseases. Expert Opin Biol Ther 2004;4:1073-91.
- Qian C, Prieto J. Gene therapy of cancer: Induction of anti-tumor immunity. Cell Mol Immunol 2004;1:105-11.
- 24. Benaim E, Sorrentino BP. Gene therapy in pediatric oncology. Invest New Drugs 1996;14:87-99.
- 25. Qian C, Liu XY, Prieto J. Therapy of cancer by cytokines mediated by gene therapy approach. Cell Res 2006;16:182-8.
- Barzon L, Stefani AL, Pacenti M, Palù G. Versatility of gene therapy vectors through viruses. Expert Opin Biol Ther 2005;5:639-62.
- Keung EZ, Nelson PJ, Conrad C. Concise review: genetically engineered stem cell therapy targeting angiogenesis and tumor stroma in gastrointestinal malignancy. Stem Cells 2013;31:227-35.
- Lundberg C, Björklund T, Carlsson T, Jakobsson J, Hantraye P, Déglon N, *et al*. Applications of lentiviral vectors for biology and gene therapy of neurological disorders. Curr Gene Ther 2008;8:461-73.
- Nanou A, Azzouz M. Gene therapy for neurodegenerative diseases based on lentiviral vectors. Prog Brain Res 2009;175:187-200.
- Braverman NE, D'Agostino MD, Maclean GE. Peroxisome biogenesis disorders: Biological, clinical and pathophysiological perspectives. Dev Disabil Res Rev 2013;17:187-96.
- Hung KL, Wang JS, Keng WT, Chen HJ, Liang JS, Ngu LH, et al. Mutational analyses on X-linked adrenoleukodystrophy reveal a novel cryptic splicing and three missense mutations in the ABCD1 gene. Pediatr Neurol 2013;49:185-90.
- Ezgu F. Recent advances in the molecular diagnosis of inborn errors of metabolism. Clin Biochem 2014;47:759-60.
- 33. Cheng SH. Gene therapy for the neurological manifestations in lysosomal storage disorders. J Lipid Res 2014;55:1827-38.
- 34. Macnab S, Whitehouse A. Progress and prospects: human artificial chromosomes. Gene Ther 2009;16:1180-8.
- Beg S, Rizwan M, Sheikh AM, Hasnain MS, Anwer K, Kohli K. Advancement in carbon nanotubes: basics, biomedical applications and toxicity. J Pharm Pharmacol 2011;63:141-63.
- Mallick K, Strydom AM. Biophilic carbon nanotubes. Colloids Surf B Biointerfaces 2013;105:310-8.
- Rastogi V, Yadav P, Bhattacharya SS, Mishra AK, Verma N, Verma A, et al. Carbon nanotubes: an emerging drug carrier for targeting cancer cells. J Drug Deliv 2014;2014:670815.

- Plusa B, Piotrowska K, Zernicka-Goetz M. Sperm entry position provides a surface marker for the first cleavage plane of the mouse zygote. Genesis 2002;32:193-8.
- Takahashi K, Okita K, Nakagawa M, Yamanaka S. Induction of pluripotent stem cells from fibroblast cultures. Nat Protoc 2007;2:3081-9.
- Yu J, Vodyanik MA, Smuga-Otto K, Antosiewicz-Bourget J, Frane JL, Tian S, *et al.* Induced pluripotent stem cell lines derived from human somatic cells. Science 2007;318:1917-20.
- Wilmut I. The first direct reprogramming of adult human fibroblasts. Cell Stem Cell 2007;1:593-4. Induction of pluripotency: from mouse to human. Cell 2007;131:834-5.
- 42. Zaehres H, Schöler HR
- 43. Wernig M, Zhao JP, Pruszak J, Hedlund E, Fu D, Soldner F, et al. Neurons derived from reprogrammed fibroblasts functionally integrate into the fetal brain and improve symptoms of rats with Parkinson's disease. Proc Natl Acad Sci U S A 2008;105:5856-61.
- Okita K, Nakagawa M, Hyenjong H, Ichisaka T, Yamanaka S. Generation of mouse induced pluripotent stem cells without viral vectors. Science 2008;322:949-53.
- 45. Yakubov E, Rechavi G, Rozenblatt S, Givol D. Reprogramming of human fibroblasts to pluripotent stem cells using mRNA of four transcription factors. Biochem Biophys Res Commun 2010;394:189-93.
- Pu J, Jiang H, Zhang B, Feng J. Redefining Parkinson's disease research using induced pluripotent stem cells. Curr Neurol Neurosci Rep 2012;12:392-8.
- 47. Bishop AE. Pulmonary epithelial stem cells. Cell Prolif 2004;37(1):89-96.
- Liu X, Driskell RR, Engelhardt JF. Airway glandular development and stem cells. Curr Top Dev Biol 2004;64:33-56.
- 49. Hoffman JA, Merrill BJ. New and renewed perspectives on embryonic stem cell pluripotency. Front Biosci 2007;12:3321-32.
- 50. Thapar N. New frontiers in the treatment of Hirschsprung disease. J Pediatr Gastroenterol Nutr 2009;48 Suppl 2:S92-4.
- Dhaulakhandi DB, Rohilla S, Rattan KN. Neural tube defects: review of experimental evidence on stem cell therapy and newer treatment options. Fetal Diagn Ther 2010;28:72-8.
- Roomans GM. Tissue engineering and the use of stem/progenitor cells for airway epithelium repair. Eur Cell Mater 2010; 19:284-99.
- Somers A, Jean JC, Sommer CA, Omari A, Ford CC, Mills JA, *et al.* Generation of transgene-free lung disease-specific human induced pluripotent stem cells using a single excisable lentiviral stem cell cassette. Stem Cells 2010;28:1728-40.
- 54. Hotta R, Natarajan D, Burns AJ, Thapar N. Stem cells for GI motility disorders. Curr Opin Pharmacol 2011;11:617-23.
- 55. Bernstein HS, Srivastava D. Stem cell therapy for cardiac disease. Pediatr Res 2012;71:491-9.
- 56. Files MD, Boucek RJ. 'Shovel-Ready' applications of stem cell advances for pediatric heart disease. Curr Opin Pediatr 2012;24:577-83.
- Wong AP, Bear CE, Chin S, Pasceri P, Thompson TO, Huan LJ, *et al.* Directed differentiation of human pluripotent stem cells into mature airway epithelia expressing functional CFTR protein. Nat Biotechnol 2012;30:876-82.
- Green MD, Huang SX, Snoeck HW. Stem cells of the respiratory system: from identification to differentiation into functional epithelium. Bioessays 2013;35:261-70.
- 59. Maher KO, Xu C. Marching towards regenerative cardiac therapy with human pluripotent stem cells. Discov Med 2013;15:349-56.
- 60. Moodley Y, Thompson P, Warburton D. Stem cells: a recapitulation of development. Respirology 2013;18:1167-76.
- 61. Nakamura K, Hirano K, Wu SM. iPS cell modeling of cardiometabolic diseases. J Cardiovasc Transl Res 2013;6:46-53.
- Saadai P, Wang A, Nout YS, Downing TL, Lofberg K, Beattie MS, et al. Human induced pluripotent stem cell-derived neural crest stem cells integrate into the injured spinal cord in the fetal lamb model of myelomeningocele. J Pediatr Surg 2013;48:158-63.
- Barad L, Schick R, Zeevi-Levin N, Itskovitz-Eldor J, Binah O. Human embryonic stem cells vs human induced pluripotent stem cells for cardiac repair. Can J Cardiol 2014;30:1279-87.
- 64. Chen G, Li S, Karakikes I, Ren L, Chow MZ, Chopra A, et al. Phospholamban as a crucial determinant of the inotropic response of human pluripotent stem cell-derived ventricular cardiomyocytes and engineered 3D tissue constructs. Circ Arrhythm Electrophysiol 2014.
- Citro L, Naidu S, Hassan F, Kuppusamy ML, Kuppusamy P, Angelos MG, et al. Comparison of human induced pluripotent stem-

cell derived cardiomyocytes with human mesenchymal stem cells following acute myocardial infarction. PLoS One 2014;9:e116281.

- Dianat N, Dubois-Pot-Schneider H, Steichen C, Desterke C, Leclerc P, Raveux A, *et al.* Generation of functional cholangiocyte-like cells from human pluripotent stem cells and HepaRG cells. Hepatology 2014;60:700-14.
- Doerr L, Thomas U, Guinot DR, Bot CT, Stoelzle-Feix S, Beckler M, et al. New easy-to-use hybrid system for extracellular potential and impedance recordings. J Lab Autom 2014.
- Drawnel FM, Boccardo S, Prummer M, Delobel F, Graff A, Weber M, et al. Disease modeling and phenotypic drug screening for diabetic cardiomyopathy using human induced pluripotent stem cells. Cell Rep 2014;9:810-21.
- Egashira T, Yuasa S, Tohyama S, Kuroda Y, Suzuki T, Seki T, et al. Patientspecific induced pluripotent stem cell models: Characterization of iPS cell-derived cardiomyocytes. Methods Mol Biol 2014.
- Firth AL, Dargitz CT, Qualls SJ, Menon T, Wright R, Singer O, *et al.* Generation of multiciliated cells in functional airway epithelia from human induced pluripotent stem cells. Proc Natl Acad Sci U S A 2014;111:E1723-30.
- Földes G, Matsa E, Kriston-Vizi J, Leja T, Amisten S, Kolker L, et al. Aberrant a-adrenergic hypertrophic response in cardiomyocytes from human induced pluripotent cells. Stem Cell Reports 2014;3:905-14.
- 72. Gomperts BN. Induction of multiciliated cells from induced pluripotent stem cells. Proc Natl Acad Sci U S A 2014;111:6120-1.
- Hetz S, Acikgoez A, Voss U, Nieber K, Holland H, Hegewald C, *et al.* In vivo transplantation of neurosphere-like bodies derived from the human postnatal and adult enteric nervous system: a pilot study. PLoS One 2014;9:e93605.
- Holmgren G, Synnergren J, Bogestål Y, Améen C, Åkesson K, Holmgren S, *et al.* Identification of novel biomarkers for doxorubicininduced toxicity in human cardiomyocytes derived from pluripotent stem cells. Toxicology 2015;328:102-11.
- Huang SX, Islam MN, O'Neill J, Hu Z, Yang YG, Chen YW, *et al.* Efficient generation of lung and airway epithelial cells from human pluripotent stem cells. Nat Biotechnol 2014;32:84-91.
- Jiang Y, Habibollah S, Tilgner K, Collin J, Barta T, Al-Aama JY, *et al.* An induced pluripotent stem cell model of hypoplastic left heart syndrome (HLHS) reveals multiple expression and functional differences in HLHS-derived cardiac myocytes. Stem Cells Transl Med 2014;3:416-23.
- Jones DK, Liu F, Vaidyanathan R, Eckhardt LL, Trudeau MC, Robertson GA. hERG 1b is critical for human cardiac repolarization. Proc Natl Acad Sci U S A 2014;111:18073-7.
- Kobayashi J, Yoshida M, Tarui S, Hirata M, Nagai Y, Kasahara S, et al. Directed differentiation of patient-specific induced pluripotent stem cells identifies the transcriptional repression and epigenetic modification of NKX2-5, HAND1, and NOTCH1 in hypoplastic left heart syndrome. PLoS One 2014;9:e102796.
- Lim SY, Sivakumaran P, Crombie DE, Dusting GJ, Pébay A, Dilley RJ. Enhancing human cardiomyocyte differentiation from induced pluripotent stem cells with trichostatin A. Methods Mol Biol 2014.
- Lu J, Wei H, Wu J, Jamil MF, Tan ML, Adenan MI, *et al.* Evaluation of the cardiotoxicity of mitragynine and its analogues using human induced pluripotent stem cell-derived cardiomyocytes. PLoS One 2014;9:e115648.
- Natarajan D, Cooper J, Choudhury S, Delalande JM, McCann C, Howe SJ, *et al*. Lentiviral labeling of mouse and human enteric nervous system stem cells for regenerative medicine studies. Neurogastroenterol Motil 2014;26:1513-8.
- Pointon A, Harmer AR, Dale IL, Abi-Gerges N, Bowes J, Pollard C, et al. Assessment of cardiomyocyte contraction in human-induced pluripotent stem cell-derived cardiomyocytes. Toxicol Sci 2014. pii: kfu312.
- Ramalingam S, Annaluru N, Kandavelou K, Chandrasegaran S. TALEN-mediated generation and genetic correction of diseasespecific human induced pluripotent stem cells. Curr Gene Ther 2014;14:461-72.
- Raval KK, Tao R, White BE, De Lange WJ, Koonce CH, Yu J, *et al.* Pompe disease results in a golgi-based glycosylation deficit in human induced pluripotent stem cell-derived cardiomyocytes. J Biol Chem 2015;290:3121-36.
- Sallam K, Kodo K, Wu JC. Modeling inherited cardiac disorders. Circ J 2014;78:784-94.

- Sargent RG, Suzuki S, Gruenert DC. Nuclease-mediated doublestrand break (DSB) enhancement of small fragment homologous recombination (SFHR) gene modification in human-induced pluripotent stem cells (hiPSCs). Methods Mol Biol 2014;1114:279-90.
- van den Heuvel NH, van Veen TA, Lim B, Jonsson MK. Lessons from the heart: Mirroring electrophysiological characteristics during cardiac development to *in vitro* differentiation of stem cell derived cardiomyocytes. J Mol Cell Cardiol 2014;67:12-25.
- Wong AP, Rossant J. Generation of Lung Epithelium from Pluripotent Stem Cells. Curr Pathobiol Rep 2013;1:137-45.
- Yang X, Pabon L, Murry CE. Engineering adolescence: maturation of human pluripotent stem cell-derived cardiomyocytes. Circ Res 2014;114:511-23.
- Ye L, Chang YH, Xiong Q, Zhang P, Zhang L, Somasundaram P, et al. Cardiac repair in a porcine model of acute myocardial infarction with human induced pluripotent stem cell-derived cardiovascular cells. Cell Stem Cell 2014;15:750-61.
- Zanella F, Sheikh F. Patient-Specific Induced Pluripotent Stem Cell Models: Generation and Characterization of Cardiac Cells. Methods Mol Biol 2014.
- Zarogoulidis P, Hohenforst-Schmidt W, Huang H, Sahpatzidou D, Freitag L, Sakkas L, *et al*. A gene therapy induced emphysema model and the protective role of stem cells. Diagn Pathol 2014 14;9:195.
- Zhang J, Qu J, Wang J. Patch clamp apply in cardiomyocytes derived from patient's iPS cells for individual anticancer therapy. Int J Clin Exp Med 2014;7:4475-8.
- Zhang L, Guo J, Zhang P, Xiong Q, Wu SC, Xia L, *et al.* Derivation and high engraftment of patient-specific cardiomyocyte sheet using induced pluripotent stem cells generated from adult cardiac fibroblast. Circ Heart Fail 2015;8:156-66.
- Zhang M, D'Aniello C, Verkerk AO, Wrobel E, Frank S, Ward-van Oostwaard D, et al. Recessive cardiac phenotypes in induced pluripotent stem cell models of Jervell and Lange-Nielsen syndrome: Disease mechanisms and pharmacological rescue. Proc Natl Acad Sci U S A 2014;111:E5383-92.
- Chen LW, Kuang F, Wei LC, Ding YX, Yung KK, Chan YS. Potential application of induced pluripotent stem cells in cell replacement therapy for Parkinson's disease. CNS Neurol Disord Drug Targets 2011;10:449-58.
- Nishimura K, Takahashi J. Therapeutic application of stem cell technology toward the treatment of Parkinson's disease. Biol Pharm Bull 2013;36:171-5.
- Koba C, Haruta M, Matsunaga Y, Matsumura K, Haga E, Sasaki Y, *et al.* Therapeutic effect of human iPS-cell-derived myeloid cells expressing IFN-ß against peritoneally disseminated cancer in xenograft models. PLoS One 2013;8:e67567.
- Wakao H, Yoshikiyo K, Koshimizu U, Furukawa T, Enomoto K, Matsunaga T, *et al.* Expansion of functional human mucosalassociated invariant T cells via reprogramming to pluripotency and redifferentiation. Cell Stem Cell 2013;12:546-58.
- 100. Musunuru K. Genome editing of human pluripotent stem cells to generate human cellular disease models. Dis Model Mech 2013;6:896-904.
- 101. Sundberg M, Bogetofte H, Lawson T, Jansson J, Smith G, Astradsson A, *et al.* Improved cell therapy protocols for Parkinson's disease based on differentiation efficiency and safety of hESC-, hiPSC-, and non-human primate iPSC-derived dopaminergic neurons. Stem Cells 2013;31:1548-62.
- Imaizumi Y, Okano H. Modeling human neurological disorders with induced pluripotent stem cells. J Neurochem 2014;129:388-99.
- 103. Makridakis M, Roubelakis MG, Vlahou A. Stem cells: insights into the secretome. Biochim Biophys Acta 2013;1834:2380-4.
- Zimmerlin L, Park TS, Zambidis ET, Donnenberg VS, Donnenberg AD. Mesenchymal stem cell secretome and regenerative therapy after cancer. Biochimie 2013;95:2235-45.
- 105. Maumus M, Jorgensen C, Noël D. Mesenchymal stem cells in regenerative medicine applied to rheumatic diseases: role of secretome and exosomes. Biochimie 2013;95:2229-34.
- 106. Drago D, Cossetti C, Iraci N, Gaude E, Musco G, Bachi A, et al. The stem cell secretome and its role in brain repair. Biochimie 2013;95:2271-85.
- Drouet V, Ruiz M, Zala D, Feyeux M, Auregan G, Cambon K, *et al.* Allele-specific silencing of mutant huntingtin in rodent brain and human stem cells. PLoS One 2014;9:e99341.

- De Coppi P, Bartsch G Jr, Siddiqui MM, Xu T, Santos CC, Perin L, *et al.* Isolation of amniotic stem cell lines with potential for therapy. Nat Biotechnol 2007;25:100-6.
- 109. Skardal A, Mack D, Kapetanovic E, Atala A, Jackson JD, Yoo J, et al. Bioprinted amniotic fluid-derived stem cells accelerate healing of large skin wounds. Stem Cells Transl Med 2012;1:792-802.
- Pozzobon M, Piccoli M, Schiavo AA, Atala A, De Coppi P. Isolation of c-Kit+ human amniotic fluid stem cells from second trimester. Methods Mol Biol 2013;1035:191-8.
- 111. Skardal A, Mack D, Atala A, Soker S. Substrate elasticity controls cell proliferation, surface marker expression and motile phenotype in amniotic fluid-derived stem cells. J Mech Behav Biomed Mater 2013;17:307-16.
- 112. Pipino C, Di Tomo P, Mandatori D, Cianci E, Lanuti P, Cutrona MB, et al. Calcium sensing receptor activation by calcimimetic R-568 in human amniotic fluid mesenchymal stem cells: correlation with osteogenic differentiation. Stem Cells Dev 2014;23:2959-71.
- 113. Xiao GY, Liu IH, Cheng CC, Chang CC, Lee YH, Cheng WT, et al. Amniotic fluid stem cells prevent follicle atresia and rescue fertility of mice with premature ovarian failure induced by chemotherapy. PLoS One 2014;9:e106538.
- 114. Ma X, Li H, Xin S, Ma Y, Ouyang T. Human amniotic fluid stem cells support undifferentiated propagation and pluripotency of human embryonic stem cell without b-FGF in a density dependent manner. Int J Clin Exp Pathol 2014;7:4661-73.
- 115. Ramachandra DL, Shaw SS, Shangaris P, Loukogeorgakis S, Guillot PV, Coppi PD, David AL. *In utero* therapy for congenital disorders using amniotic fluid stem cells. Front Pharmacol 2014;5:270.

- 116. Tajiri N, Acosta S, Portillo-Gonzales GS, Aguirre D, Reyes S, Lozano D, et al. Therapeutic outcomes of transplantation of amniotic fluidderived stem cells in experimental ischemic stroke. Front Cell Neurosci 2014 13;8:227.
- 117. Kastenberg ZJ, Odorico JS. Alternative sources of pluripotency: science, ethics, and stem cells. Transplant Rev (Orlando) 2008;22:215-22.
- Kovacs GT, Breheny SA, Dear MJ. Embryo donation at an Australian university *in-vitro* fertilisation clinic: issues and outcomes. Med J Aust 2003;178:127-9.
- 119. Alizadeh L, Omani Samani R. Using fertile couples as embryo donors: An ethical dilemma. Iran J Reprod Med 2014;12:169-74.
- Amesse LS, Srivastava G, Uddin D, Pfaff-Amesse T. Comparison of cryopreserved sperm in vaporous and liquid nitrogen. J Reprod Med 2003;48:319-24.
- 121. Crespilho AM, Nichi M, Guasti PN, Freitas-Dell'Aqua CP, Sá Filho MF, Maziero RR, *et al.* Sperm fertility and viability following 48h of refrigeration: evaluation of different extenders for the preservation of bull semen in liquid state. Anim Reprod Sci 2014;146:126-33.

© SAGEYA. This is an open access article licensed under the terms of the Creative Commons Attribution Non-Commercial License (http:// creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted, noncommercial use, distribution and reproduction in any medium, provided the work is properly cited.

Source of Support: Nil, Conflict of Interest: None declared.